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## BIOFILM FORMATOIN POTENTIAL (BFP) OF *Salmonella Spp.* ISOLATED FROM STREET VENDED READY-TO-EAT FRIED GRASSHOPPER: A THREAT TO FOOD SAFETY

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#### ABSTRACT

Bacterial biofilms are serious global public health problem due to their antibiotics tolerance capabilities, resistance to host defence mechanisms and other external stresses which consequently contributes to their chronic infections. RTE (ready-to eat) grasshopper is much consumed by women and children and its marketability is overtaking that of meat in many parts of Northeastern Nigeria and can serve as reservoir for biofilm forming pathogenic bacteria. This study aimed to detect the presence of *Salmonella sp.* from exposed fried grasshopper vended in Gombe metropolis and to assess their biofilm formation potential (BFP). A total of 250 samples of exposed RTE fried grasshopper were purchased from different locations of Gombe metropolis in sterile containers. They were homogenized in Rappaport Vassiliadis *Salmonella* enrichment broth and spread plated on selective *Salmonella-Shigella* agar. Positive samples were Gram stained and further subjected to biochemical identifications. The confirmed isolates were further assessed for BFP using Congo red agar assay technique. Of the 250 samples obtained, 36% (n=90) were positive for *Salmonella sp.* out of which 77.8% (n=70) had biofilm formation potential. Presence of *Salmonella sp.* with BFP in foods is a serious concern that can lead to outbreaks as the biofilm may be protection for their persistence. Relevant authorities should take serious measures on the strict adherence to proper food hygiene practices to save the consumers from outbreaks due to the organisms.

**Keywords:** *Salmonella sp.*, grasshopper, ready-to-eat, biofilm formation potential, Gombe metropolis.

#### Introduction

The usage of insects for human consumption as a source of nutrient and to combat famine has been frequently practiced globally, mainly in Africa (including but not limited to Nigeria), Australia, America, Thailand and other countries in the Asia (Milanović et al., 2018). There are indeed various promising aspects associated with the consumption of insects as they are universally characterized by nutritionally positive profiles for the presence of high-quality amino acids and proteins, vitamins, good lipids, fiber and minerals. They can also breed as easily and soon as possible, and are capable of emitting ammonia and

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greenhouse gases lower than traditional livestock do (Schlüter *et al.*, 2017; Milanović *et al.*, 2018). As a result of their numerous nutritional, environmental and social benefits, edible insects are recognized as the “food of the future” and are equally categorized as novel foods by Regulation (EU) No 2015/2283 of the European Parliament and Council (Milanović *et al.*, 2018).

Grasshoppers (*Ruspolia differens*) are among the edible insects already commercialized as foods in the countries of the European Union (EU) (Schlüter *et al.*, 2017) including Africa that have not been fully investigated for the presence of pathogenic bacteria of relevance or as a whole, potential microbial pathogens (Ali *et al.*, 2010; Klunder *et al.*, 2012; Stoops *et al.*, 2016; Garofalo *et al.*, 2017).

Foodborne infections and/or intoxications occur due to ingestion of food that is polluted by pathogens, their toxins or poisons, that happens at any stage of food processing or handling from in the food chain. Foodborne diseases or contaminations mostly happen because of eating such polluted, ruined, poisonous or toxic foods. Some of the manifestations associated with food contamination include queasiness, heaving and loose bowels (Gupta *et al.*, 2019).

*Salmonella* spp. is an important causative agent of foodborne diseases responsible for more than a million cases annually (Beshiru *et al.*, 2017). It is considered the most important foodborne pathogen capable of causing severe infections in humans and global economic losses. Its presence is regularly monitored in various steps of the food chain, and particularly among the finished raw and ready-to-eat (RTE) products, as among the criteria for consumer safety which represents a very important tool for the implementation of efficient systems for food safety (Tirziu *et al.*, 2020).

Salmonellosis has become a major global public health concern and emerged as a significant foodborne disease leading to a significant economic and public health burden. Although direct contact with infected animals was also reported for acquiring Salmonellosis, it is acquired generally via exposure to food, (Majowicz *et al.*, 2010; Kumar *et al.*, 2019).

Biofilm is a stationary microbial community that colonizes and grows on surfaces of substances including foods. This is achieved by the self-production of extracellular polymeric substances (EPS) that help them become firmly attached; thereby causing infections that could only be treated by its removal which in turn leads to unaffordable treatments and mental-illness to the individuals. Bacterial biofilms pose a serious health concern worldwide due to their capability for tolerating host defence systems, antibiotics and other external stresses. It thus contributes to persistence with chronic infections (Høiby *et al.*, 2011; Sharma *et al.*, 2019). Biofilm comprises of the crammed bacterial population by extra-cellular matrix (ECM) which possesses bacterial secreted polymers such as exopolysaccharides (EPS), extracellular DNA (e-DNA), proteins and amyloidogenic proteins (Whitchurch *et al.*, 2002).

The protections that biofilms provide to microorganisms within it are not only limited to against nutrients scarcity, altered pH, mechanical and shear forces and osmolarity, but also block the access of host's immunity and antibiotics to the bacterial communities. Thus, the matrix of biofilm provides the additional power of bacterial resistance that renders them tolerable to harsh conditions and antibiotics. This brings about the emergence of infections of the bad bugs such as multidrug resistant (MDR), extensively drug resistant (XDR) and

totally drug resistant bacteria (McCarty *et al.*, 2012; Sharma *et al.*, 2019).

An investigation conducted on degutted, washed, spiced, roasted, and sundried *Bunaea alcinoe* (grasshopper) hatchlings revealed the presence of *Pseudomonas* sp and *Proteus* sp. In expansion, there is a danger of microbial contamination of the bug item because of the outside and the street side climates where they are callously showcased just as legitimate food handling practices and information by vendors were absent (Mugo, 2020). Therefore, this study aimed to determine the presence of *Salmonella* species from uncovered RTE fried grasshopper with the objective of evaluating their biofilm formation capabilities.

## Materials and Methods

### Sample Collection and Processing

A total of 250 samples of exposed grasshoppers were purchased randomly from different vendors within Gombe metropolis and aseptically introduced into sterile sampling bottles. They were then transported to the laboratory at the Centre of Excellence for Food Safety Research Faculty of Food Science and Technology, Universiti Putra Malaysia (UPM) for analysis. A 25 g of fried grasshopper sample was added to 225 ml Rappaport Vassiliadis *Salmonella* enrichment broth and the mixture was homogenized in a stomacher bag for five minutes. One ml aliquot was pipetted into a Petri dish containing salmonella-shigella agar and incubated at 37 °C for 24 hours. Upon incubation, the black colonies observed on SS agar are typical of *Salmonella* species.

### Isolation and Identification of the Isolates

The dark colonies obtained on the SS agar were isolated using pure culture isolation technique on nutrient agar and then subjected to Gram staining technique and biochemical tests such as urease, citrate utilization, indole production, glucose fermentation, motility, mannitol fermentation and growth on triple sugar iron (TSI) agar to confirm their identities.

### Standardization of Inoculum for Biofilm Assay

The confirmed isolates were further subcultured on nutrient agar (NA) and incubated for overnight. Thereafter, a portion of the colony was emulsified into sterile buffered peptone water (BPW) to match the 0.5 MacFarland turbidity standard, equivalent to  $1.5 \times 10^8$  CFU/ml.

### Biofilm formation Potential Assay

The assay for biofilm formation potential (BFP) was conducted using phenotypic expression of colonies in Congo Red Agar (CRA). The CRA was prepared by dissolving 37 g Brain Heart Infusion (BHI) agar (Titan Biotech Ltd, INDIA), 36 g sucrose and 0.8 g congo red (BDH Ltd, India) in one liter of distilled water. The solution was sterilized by autoclaving at 121°C for 15 minutes and dispensed in plates. Upon solidification, the medium was inoculated with the *Salmonella* isolates and incubated at 37 °C for 18 hours. Isolates with the potential for biofilm formation showed black colonies while those without the BFP remained red.

## Results

Figure 1 depicts that of the 250 exposed fried RTE grasshopper samples, 90 were found to be positive following selective isolation on SSA and Gram staining and biochemical tests. This accounts for the 36% of the total samples. *Shigella* species had an occurrence of 145 samples (58%) while other unidentified organisms accounted for the 6% (n=15) of the samples. The same results are also presented in figure 1 below.

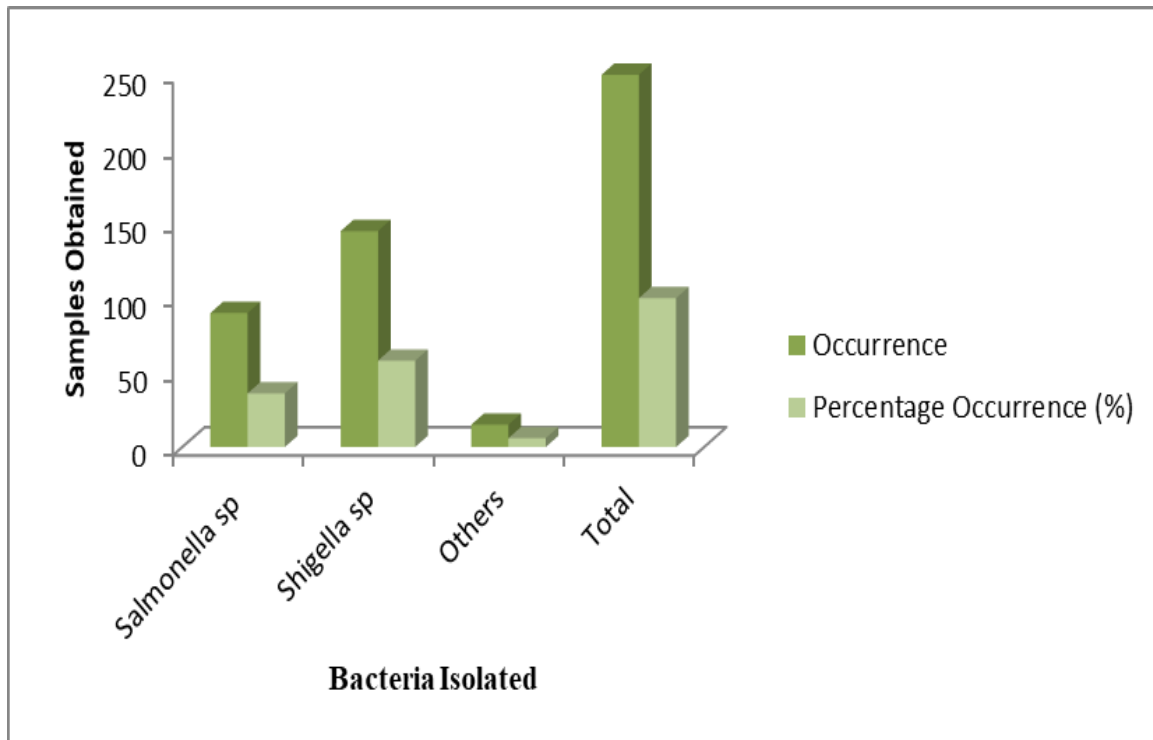


Fig. 1: Occurrence of *Salmonella sp.* from RTE samples of exposed Grasshopper

Out of the 90 confirmed isolates obtained, it can be observed that 70 were found to have the potential of producing biofilm (Table 1) while the remaining 20 of *Salmonella sp.* isolates obtained were negative for biofilm formation potential (BFP) assay. This brings about the percentages of both the positive and negative isolates as 77.7% and 22.2% respectively. The BFP positive isolates are identified based on the appearance of their colonies on Congo red agar (CRA). They produced black colonies on the CRA, an indication of their potential to produce biofilm (Fig. 2).

Table 1: *Salmonella sp.* with BFP isolated from exposed RTE grasshopper

Isolate	Positive for BFP	Negative for BFP	% positive	% negative	Total
<i>Salmonella sp</i>	70	20	77.8	22.2	90



Fig 2: *Salmonella sp.* isolates with biofilm formation potential. The dark swarming colonies indicate BPF and motility, a typical of motile, biofilm-forming *Salmonella*.

## Discussion

Insects are considered to be a ‘food for the future’ as a result of their positive nutritional characteristics and low environmental impact. The safety associated with their consumption is a growing concern. Such a concern may be, but not limited to their link with contaminating chemical and biological agents. The potential for the pathogenic and toxigenic microbes’ presence in ready-to-eat edible insects is among the main biological hazards associated with edible insects (Garofalo *et al.*, 2019). Entomophagy as the consumption of edible insects and has been globally practiced for long by humans (Belluco *et al.*, 2015). It has been investigated that approximately 2000 insect species or more are consumed, with the main consumers in Asia, Africa, Latin America (e.g., Mexico), and Australia. In some of these regions, insects are important and typical part of the diets of humans, with their importantly cultural and gastronomical aspects (van Huis *et al.*, 2016).

However, there are issues associated with the safety of consuming edible insects. In 2015, a Scientific Opinion was issued by the European Food Safety Authority (EFSA) that is concerned with the profile of the risk in relation with the production and consumption of insects as food and feed (EFSA, 2015). The opinion explored the possible issues of safety such as the biological, chemical, and environmental; that are associated with farmed, processed, and non-processed insects along the whole production chain. It was concluded that many factors could have an effect on the edible insects’ safety, including the method of production, the substrate, the harvest stage, the species of insect, insect developmental stage and the insect processing. The opinion also highlighted the lack of scientific studies on the possible hazards when insects are used as food and feed, together with a paucity of

systematically collected data on animal and human consumption of insects. In addition, there is little or no data on the presence of foodborne potential pathogens like *Salmonella sp* that have the capability of biofilm production isolated from exposed RTE grasshopper vended in the study area. Thus, this study serves as a short communication to enlighten the concerned populace on the potential risks associated with the organisms, which may eventually leads to outbreak that may prove difficult to contain.

Several studies have highlighted a typical microbial profile for grasshoppers, both fresh and processed (boiled and dried), collected from different producers various parts of the world with the prevalence of bacteria and fungi (Garofalo *et al.*, 2019). The microbiota in the grasshopper was characterized by the dominating of LAB (lactic acid bacteria), mainly of the genera *Pediococcus*, *Weissella*, *Enterococcus* and *Lactococcus*, and species belonging to the Enterobacteriaceae, such as *Klebsiella/Enterobacter spp*. However, *Salmonella sp.* was not detected. *Staphylococcus sp.* and *Bacillus sp.* were prevalent in grasshopper samples in Thailand (Milanović *et al.*, 2018) but with no traces of *Salmonella sp.*, hence the uniqueness of this study.

Detection of *Salmonella sp.* in grasshopper has been rare as Stoop *et al.*, (2016) also was able to detect only *Enterobacter sp.*, *Klebsiella sp.* and *Yersinia sp.* among the members of Enterobacteriaceae. Similar research conducted by Grabowski and Klein (2017) on fried grasshopper also did not reveal the presence of *Salmonella sp.*, *Staphylococcus aureus*, *E. coli* and *Listeria monocytogenes*. The detection of Salmonellae in this study is therefore is imperative to the food safety of the populace, as consumption of the RTE grasshopper in northern Nigeria and Gombe metropolis has been on the increase. No data so far has been published to the knowledge of the authors on the detection of *Salmonella sp.* from fried grasshopper in Nigeria and elsewhere where the grasshopper is grossly consumed.

About 80% of chronic and recurrent human infections due to microorganisms are due to bacterial biofilms. The cells of microorganisms within a biofilm have been shown to have 10–1000 times more resistant to mechanical stress and antibiotics than their planktonic counterparts. The uptake of resistance gene by the horizontal gene transfer is among mechanisms of resistance to antibiotics by bacterial. Moreover, increased genetic competence, high cell density, accumulation of genetic elements or resistance genes uptake and increased genetic competence are compatible conditions provided by biofilms for their survival (Mah, 2012).

*Salmonella sp.* possesses the capability of biofilm formation on several surfaces, and this favours its survival in hostile environments such as fried foods, utensils and slaughter houses (Borges *et al.*, 2018). The presence of biofilm formation potential of *Salmonella sp.* on various surfaces has been reported by several studies (Agarwal *et al.*, 2006; Tondo *et al.*, 2010; Steenackers *et al.*, 2012; Borges *et al.*, 2018). In this study, the report of the BFP of 77.8% of the *Salmonella sp.* isolated from exposed vended RTE grasshopper is an indication of risks associated with the consumption of the grasshopper. This if left unchecked can eventually lead to outbreak due to this organism and because of the BFP associated with it, which may consequently lead to loss lives and properties.

## Conclusion

This study provides an insight on the risks associated with the exposed RTE fried grasshopper vended in the streets of Gombe metropolis, Gombe State Nigeria. A substantial number of the samples were found to harbor *Salmonella sp.* out of which approximately 78% were of biofilm formation potential (BFP). This may consequently confer on them the ability

to resist antibiotics and other mechanical stresses, thereby given them a strong chance for survival.

## RECOMMENDATION

This study is a short communication; highlighting the potential presence of *Salmonella sp.* from fried grasshopper. More studies are therefore needed for the presence of antibiotic resistance as well as the use of various methods for the assessment of biofilm formation. Further studies are also encouraged to ascertain the cause of the presence of *Salmonella sp.* in the street vended exposed fried grasshopper so as to avoid imminent outbreaks due to this organism for the avoidance of loss of lives and properties due to the organism.

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## CONFLICT OF INTEREST

There was not a conflict of interest observed among the authors during the conduct of the study

## References

- Agarwal, R.K., Singh, S., Bhilegaonkar, K.N.; Singh, V.P. (2006) Optimization of microtitre plate assay for the testing of biofilm formation ability in different *Salmonella* serotypes. *International Food Research Journal*, 18: 1493-1498.
- Ali, A., Mohamadou, B. A., Saidou, C., Aoudou, Y., & Tchiegang, C. (2010). Physico-chemical properties and safety of grasshoppers, important contributors to food security in the far north region of Cameroon. *Res J Anim Sci*, 4, 108-111.
- Belluco, S., Losasso, C., Maggioletti, M., Alonzi, C., Ricci, A., & Paoletti, M. G. (2015). Edible insects: a food security solution or a food safety concern?. *Animal Frontiers*, 5(2), 25-30.
- Beshiru, A., Igbinsosa, I. H., & Igbinsosa, E. O. (2018). Biofilm formation and potential virulence factors of *Salmonella* strains isolated from ready-to-eat shrimps. *PloS one*, 13(9), 1-22.
- Borges, K. A., Furian, T. Q., de Souza, S. N., Menezes, R., de Lima, D. A., Fortes, F. B. B., ... & Nascimento, V. P. (2018). Biofilm formation by *Salmonella* Enteritidis and *Salmonella* Typhimurium isolated from avian sources is partially related with their in vivo pathogenicity. *Microbial Pathogenesis*, 118, 238-241.

- Garofalo, C., Osimani, A., Milanović, V., Taccari, M., Cardinali, F., Aquilanti, L., ... & Clementi, F. (2017). The microbiota of marketed processed edible insects as revealed by high-throughput sequencing. *Food Microbiology*, 62, 15-22.
- EFSA Scientific Committee (2015). Scientific opinion on a risk profile related to production and consumption of insects as food and feed. *EFSA J.* 13(10), 4257.
- Garofalo, C., Milanović, V., Cardinali, F., Aquilanti, L., Clementi, F., & Osimani, A. (2019). Current knowledge on the microbiota of edible insects intended for human consumption: A state-of-the-art review. *Food Research International*, 125, 108527.
- Grabowski, N. T., & Klein, G. (2017). Microbiology of processed edible insect products—Results of a preliminary survey. *International Journal of Food Microbiology*, 243, 103-107.
- Gupta, R., Gupta, A., Singh, R. P., Singh, P. K., & Singh, R. L. (2019). Food Allergies. In *Food Safety and Human Health* (pp. 99-125). Academic Press.
- Høiby, N., Ciofu, O., Johansen, H. K., Song, Z. J., Moser, C., Jensen, P. Ø., ... & Bjarnsholt, T. (2011). The clinical impact of bacterial biofilms. *International Journal of Oral Science*, 3(2), 55-65.
- van Huis, A. (2016). Edible insects are the future?. *Proceedings of the Nutrition Society*, 75(3), 294-305.
- Klunder, H. C., Wolkers-Rooijackers, J., Korpela, J. M., & Nout, M. R. (2012). Microbiological aspects of processing and storage of edible insects. *Food Control*, 26(2), 628-631.
- Kumar, Y., Singh, V., Kumar, G., Gupta, N. K., & Tahlan, A. K. (2019). Serovar diversity of Salmonella among poultry. *The Indian journal of Medical Research*, 150(1), 92.
- Majowicz, S. E., Musto, J., Scallan, E., Angulo, F. J., Kirk, M., O'Brien, S. J., ... & International Collaboration on Enteric Disease “Burden of Illness” Studies. (2010). The global burden of nontyphoidal Salmonella gastroenteritis. *Clinical infectious diseases*, 50(6), 882-889.
- Mah T-F. (2012). Biofilm-specific antibiotic resistance. *Future Microbiol.* 2012;7: 1061–72.
- McCarty, S. M., Cochrane, C. A., Clegg, P. D., & Percival, S. L. (2012). The role of endogenous and exogenous enzymes in chronic wounds: a focus on the implications of aberrant levels of both host and bacterial proteases in wound healing. *Wound Repair and Regeneration*, 20(2), 125-136.
- Milanović, V., Osimani, A., Roncolini, A., Garofalo, C., Aquilanti, L., Pasquini, M., ... & Clementi, F. (2018). Investigation of the dominant microbiota in ready-to-eat



grasshoppers and mealworms and quantification of carbapenem resistance genes by qPCR. *Frontiers in Microbiology*, 9, 3036.

**Mugo, L. W. (2020).** *Hygiene Practices Of Vendors And Quality Of Grasshopper (Ruspolia Differens) Products Sold In Open Markets Of Uganda* (Doctoral dissertation, University of Nairobi).

**Schlüter, O., Rumpold, B., Holzhauser, T., Roth, A., Vogel, R. F., Quasigroch, W., ... & Engel, K. H. (2017).** Safety aspects of the production of foods and food ingredients from insects. *Molecular Nutrition & Food Research*, 61(6), 1600520.

**Sharma, D., Misba, L., & Khan, A. U. (2019).** Antibiotics versus biofilm: an emerging battleground in microbial communities. *Antimicrobial Resistance & Infection Control*, 8(1), 1-10.

**Steenackers, H., Hermans, K., Vanderleyden, J.; De Keersmaecker, S.C.J. (2012)** Salmonella biofilms: an overview on occurrence, structure, regulation and eradication. *Food Research International*, 45: 502-531

**Stoops, J., Crauwels, S., Waud, M., Claes, J., Lievens, B., & Van Campenhout, L. (2016).** Microbial community assessment of mealworm larvae (*Tenebrio molitor*) and grasshoppers (*Locusta migratoria migratorioides*) sold for human consumption. *Food Microbiology*, 53, 122-127.

**Tîrziu, E., Bărbălan, G., Morar, A., Herman, V., Cristina, R. T., & Imre, K. (2020).** Occurrence and antimicrobial susceptibility profile of *Salmonella* spp. in raw and ready-to-eat foods and *Campylobacter* spp. in retail raw chicken meat in Transylvania, Romania. *Foodborne Pathogens and Disease*, 17(8), 479-484.

**Tondo, E.C., Machado, T.R.M., Malheiros, P.S., Padrão, D.K., De Carvalho, A.L Brandelli A. (2010)** Adhesion and biocides inactivation of *Salmonella* on stainless steel and polyethylene. *Brazilian Journal of Microbiology*, 41: 1027-1037.

**Whitchurch, C. B., Tolker-Nielsen, T., Ragas, P. C., & Mattick, J. S. (2002).** Extracellular DNA required for bacterial biofilm formation. *Science*, 295(5559), 1487-1487.